

**Original Article****Effects of Naphthalene on Plasma Cortisol and Thyroid Levels in Immature and Mature Female Klunzingeri Mulet, *Liza klunzingeri***Zahra Yarahmadi<sup>1</sup>, Abdol Ali Movahedinia<sup>1</sup>, Sara Rastgar<sup>1</sup>, Rashid Alijani Ardeshir<sup>\*1</sup>

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**ABSTARCT**

**Background:** Polycyclic aromatic hydrocarbons (PAHs) such as naphthalene (NAP) are organic pollutants that have spread widely in littoral marine ecosystems. We aimed to study the effect of acute and prolonged exposure to naphthalene (NAP) on plasma cortisol and thyroid levels in mature and immature *Liza klunzingeri* mulet.

**Methods:** In acute stress experiment, the treatment group received 2 $\mu$ l g<sup>-1</sup> sunflower oil containing NAP (50 mg kg<sup>-1</sup>) intraperitoneally and the controls were injected with sunflower oil alone (2 $\mu$ l g<sup>-1</sup>). Blood samples were obtained from both groups after 3 h. In prolonged stress experiment, 10  $\mu$ l g<sup>-1</sup> of coconut oil containing NAP (50mg kg<sup>-1</sup>) was implanted and blood samples were obtained 72 h after injection.

**Results:** Both the acute and prolonged exposure induced significant increase in cortisol and a significant decrease ( $P<0.05$ ) in thyroid hormone T4 levels. Thyroid T3 hormone levels only decreased significantly ( $P<0.05$ ) after prolonged exposure. Acute exposure resulted in significant decrease ( $P<0.05$ ) in T3/T4 ratio only in immature fish. On the other hand, prolonged stress increased T3/T4 ratio in immature fish.

**Conclusion:** Changes in the plasma levels of these hormones indicate low physiological capacity and survival potential of fish in waters polluted with naphthalene.

**Keywords:** Hydrocortisone, Naphthalenes, Polycyclic Aromatic Hydrocarbons, Thyroid Hormones, Triiodothyronine.

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**INTRODUCTION**

Polycyclic aromatic hydrocarbons (PAHs) such as naphthalene (NAP) are organic pollutants spread widely in littoral marine ecosystems. These compounds commonly enter the water from the air, oil leaks as well as from industrial and domestic wastes [1]. PAHs are highly toxic to biological systems, and have considerable mutagenic and carcinogenic potential [2]. Furthermore, by induction of changes in the endocrine system, they may also affect physiological processes, which consequently disrupt essential processes like reproduction, metabolism, and the response to stress [3].

PAHs, as the ligands of aryl hydrocarbon receptors (AhRs), stimulate P450 1A cytochrome system, which consequently activate an enzymatic system responsible for removal of pollutants [4]. On the other hand, PAHs produce

some intermediate compounds that interrupt the production and secretion of circulating hormones such as cortisol and thyroid hormones [5]. These hormones play important roles in the responses of most animals such as fish to stress [5, 6]. Thyroid hormones (T<sub>3</sub> and T<sub>4</sub>) and cortisol secretion are under the control of the hypothalamo-pituitary-thyroid and hypothalamo-pituitary-adrenal systems [7, 8]. Thyroid hormones have crucial role in regulating the growth and balance of the hydro-mineral conditions of body fluids [9], while cortisol has a major role in regulating metabolic energy, stress responses, and immune system function [10]. Thyroid hormones and cortisol influence the metabolism of hydrocarbons [1]. Besides, they are used, in fish, as biomarkers [5]. Plasma levels of T<sub>3</sub>, and T<sub>4</sub>, and T<sub>3</sub>/T<sub>4</sub> ratio can be used as suitable indices for estimating the metabolism state with respect to growth, rate of protein

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synthesis, and oxygen consumed by tissues [5, 11, 12].

In This research, levels of thyroid and cortisol hormones were used as biomarkers to evaluate chemical stress induced by NAP before and during maturation of mullet (*Liza klunzingeri*) a fish found worldwide in coastal temperate and tropical waters.

## MATERIALS AND METHODS

Live female mullet fish ( $97 \pm 3$  g) were caught (November 2013) by trolling near Musa creek in the northern part of the Persian Gulf (Iran) and were transferred to the Imam Khomeini Fish Research Center. To adapt to the laboratory light and temperature conditions, they were kept in separate tanks (150 liter) for two weeks. To study the acute and prolonged effects of NAP exposure, two experimental protocols (injection and implantation of NAP dissolved in vegetable oil) were designed. Dosage selection was based upon published studies that evaluated the toxicological effects of PAHs on fish [13-15]. To study the acute effects of NAP, 70 fish were divided into the control and treatment groups. Under anesthesia (0.2% 2-phenoxyethanol), the fish were weighed and treated with sunflower oil ( $2\mu\text{l g}^{-1}$ ) containing NAP ( $50\text{ mg kg}^{-1}$ ) through peritoneal injection. The control group received sunflower oil alone. Three hours after injection, blood and gonad samples were taken. The procedure in the second experiment was the same as the first one except that the treatment group received a peritoneal implant of coconut oil ( $10\mu\text{l g}^{-1}$ ) containing NAP ( $50\text{ mg kg}^{-1}$ ). Blood and gonad samples were taken 72 h after NAP implantation.

Blood samples were obtained from the caudal vein using a heparinized syringe (2 ml), and were centrifuged (Hettich-D7200, Tuttlingen, Germany) at 6000 rpm for 6 min. The plasma was then rapidly frozen in liquid nitrogen, and stored at  $-80\text{ }^{\circ}\text{C}$  for further analysis. To determine the sexual phase of the fish, the gonads were removed and fixed in Bouin's solution for 48 h. Then, samples were transferred to ethanol (70%) prior to histological studies. After histological preparation, paraffin blocks were cut ( $5\text{ }\mu\text{m}$ ) and stained (hematoxiline-eosin). In the light microscope the previtellogenic and early vitellogenenic stages were identified (Olympus BH-Z- Japan) with Dinolite Digital Microscope software (AM211, ver. 3, AnMo Electronics Corp, Taiwan) [16].

## Plasma Hormone Measurements

The amounts of cortisol,  $T_3$ , and  $T_4$  hormones were measured by ELISA technique, according to the instructions of the manufacturer of a commercial kit (DIMETRA, Italy) [5, 17]. Absorbance in each well was measured at 450 nm in a microplate reader (IMMUNO-MINI; Nalge Nunc International) for hormone analysis. Samples from the fish with mature sexual oocyte and those with polygonal immature oocyte were used in the study.

## Statistical Analysis

To compare cortisol,  $T_3$  and  $T_4$  hormones levels, as well as  $T_3/T_4$  ratio in control and treatment groups at two sexual phases, two-way ANOVA was used. For multiple comparison the Student-Newman-Keuls test was used. The confidence coefficient was 95% ( $P<0.05$ ). Moreover, Sigma plots (Systat Software, ver. 11, Inc., CA, USA) were used for analyzing the data and drawing the resulting diagrams.

## Ethical Considerations

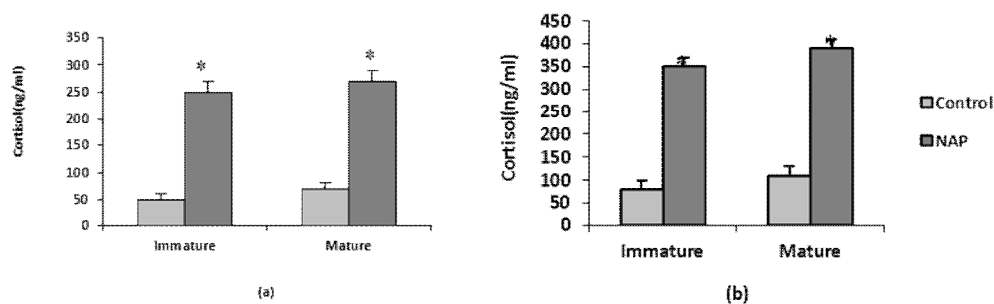
This research was performed according to convention of animal rights (approved by the Ethics Committee of Khoramshahr University of Marine Science and Technology). We tried to use animals (fish) without causing them unnecessary suffering if it could be avoided.

## RESULTS

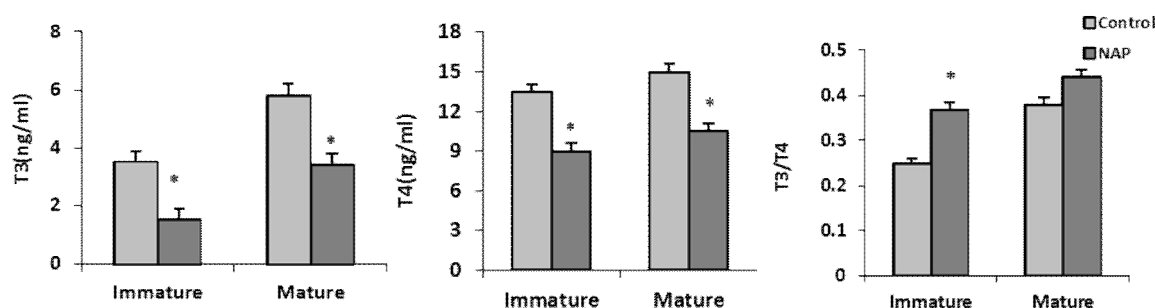
No abnormal behavior was observed in swimming, movements, and the rate of mortality during the experiments. Plasma cortisol levels were significantly increased ( $P<0.05$ ) in acute (3 h) and prolonged (72 h) exposure to NAP in mature and immature fish groups as compared to control group (Figure 1).

During the short-term exposure to NAP, the level of  $T_3$  did not change, however in both mature and immature fish,  $T_4$  levels significantly decreased ( $P<0.05$ ). The ratio of  $T_3/T_4$  showed significant decrease ( $P<0.05$ ) in immature fish (Figure 2).

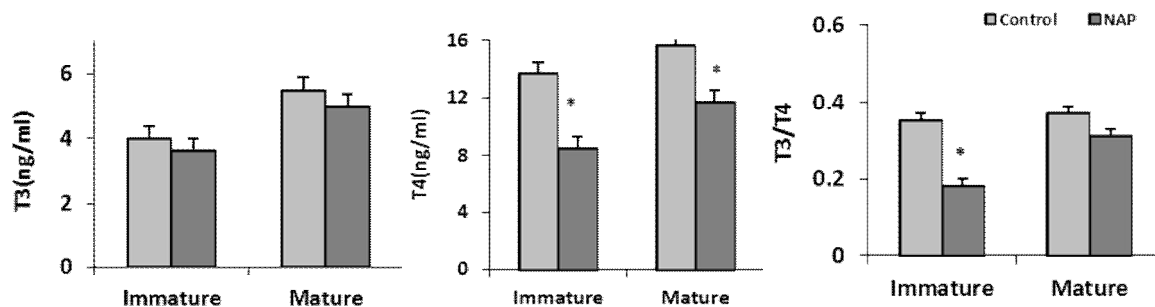
During prolonged exposure to NAP,  $T_3$  and  $T_4$  hormone levels reduced significantly ( $P<0.05$ ) in mature and immature animals. Unlike to the short-term exposure, the ratio of  $T_3/T_4$  showed a significant change (increase) ( $P<0.05$ ) in immature fish (Figure 3).



**Figure 1.** Effects of naphthalene treatment on the response of plasma cortisol levels in short term (a) and long term (b) exposure. Star symbol indicates significant difference in comparison with the control group ( $P < 0.05$ ).



**Figure 2.** Effects of short-term exposure (3h) to naphthalene on the response of plasma T3, T4 and T3/T4. Star symbol indicates significant difference.



**Figure 3.** Effects of long-term exposure (72 h) to naphthalene on the response of plasma T3, T4 and T3/T4. Star symbol indicates significant difference.

## DISCUSSION

Acute and prolonged exposure to NAP induces increase of plasma cortisol in both mature and immature mullets. These results are similar to those reported by Thomas et al. [18], Kennedy and Farrell [19], and Tintos et al. [20] on *Mugil cephalus*, *Clupea harengus*, and *Oncorhynchus mykiss* respectively. NAP activates AhRs [21], although the mechanism of action of AhRs is not fully understood but activation of these receptors affect hypothalamo-

pituitary-adrenal axis, probably increases the expression of corticotropin releasing factor (CRF) gene [22] and consequently, increases the secretion of ACTH and cortisol. On the contrary, Teles et al. [6] reported that PAH resulted in decreased plasma cortisol level but they concluded that it could be the result of hypothalamo-pituitary-adrenal axis inefficiency or loss of mitochondrial cortisol-synthesizing enzymes.

Thyroid hormones are important in growth, metabolism, and homeostasis. Secretion

of thyrotropin-releasing hormone from hypothalamus releases thyroid-stimulating hormone (TSH) from pituitary gland into the plasma, which stimulates the release of thyroid hormones. The main difference between T<sub>3</sub> and T<sub>4</sub> is their ability to bond to the receptors. T<sub>3</sub> bonds to receptors ten times more than that T<sub>4</sub>; therefore, it is considered as the main active biological form [5]. Acute exposure to NAP decreased plasma levels of T<sub>4</sub> in mature and immature fish. However, in prolonged exposure, both T<sub>3</sub> and T<sub>4</sub> levels were decreased in mature and immature animals. Changes in T<sub>3</sub>/T<sub>4</sub> ratio were just observed in the immature fish in each experiment, which was due to their greater sensitivity. Organic pollutants may decrease thyroid hormones production and change their circulating concentrations [6, 23, 24]. However, the exact mechanism of effect of these pollutants is not clear.

Alkindi et al. [25] observed that petroleum hydrocarbons might decrease plasma levels of thyroid hormones, and increase plasma cortisol, in flounder fish. Stephens et al. [26] reported similar results in *O. mykiss*. Decreased thyroid hormones and T<sub>3</sub>/T<sub>4</sub> ratio variations in regions polluted with crude oil were reported earlier [6]. Increased deiodination, biliary excretion of thyroid hormones [12], negative feedback in the hypothalamo-pituitary-thyroid axis [25], and disruption in the function of the hepatic 5'-monodeiodinase [27] would decrease T<sub>3</sub> and T<sub>4</sub> in plasma, and change T<sub>3</sub>/T<sub>4</sub> ratios. Furthermore, NAP might affect CYP1A1 and AhRs gene expression [4]. Expression of these genes may inhibit the expression of genes related to thyroid hormones receptors and even the genes responsible for thyroid hormones synthesis [5].

## CONCLUSION

It seems that acute and prolonged exposure to NAP increase plasma cortisol level and decrease thyroid hormones concentrations. These changes were more intense in immature fish. It appears that in female fish, immature ovary is more sensitive to pollutants. Considering the key role of thyroid hormones and cortisol in osmotic regulation and metabolism, changes in plasma levels of these hormones may cause low physiological capacity and survival potential of fish in polluted environment.

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## REFERENCE

- Hontela A, Dumont P, Duclos D, Fortin R. Endocrine and metabolic dysfunction in yellow perch, *Perca flavescens*, exposed to organic contaminants and heavy metals in the St. Lawrence River. *Envir Toxicol Chem* 1995;14(4):725-31.
- Pauka LM, Maceno M, Rosi SC, de Assis HCS. Embryotoxicity and biotransformation responses in zebrafish exposed to water-soluble fraction of crude oil. *Bulletin of Envir Contam Toxicol* 2011;86(4):389-93.
- Aluru N, Vijayan MM. [beta]-Naphthoflavone disrupts cortisol production and liver glucocorticoid responsiveness in rainbow trout. *Aqua Toxicol* 2004;67(3):273-85.
- Billiard SM, Hahn ME, Franks DG, Peterson RE, Bols NC, Hodson PV. Binding of polycyclic aromatic hydrocarbons (PAH) to teleost aryl hydrocarbon receptors (AHRs). *Compar Biochem Physiol Part B: Biochem Mol Biol* 2002;133(1):55-68.
- Brar NK, Waggoner C, Reyes JA, Fairey R, Kelley KM. Evidence for thyroid endocrine disruption in wild fish in San Francisco Bay, California, USA Relationships to contaminant exposures. *Aqua Toxicol* 2010;96(3):203-15.
- Teles M, Oliveira M, Pacheco M, Santos MA. Endocrine and metabolic changes in *Anguilla anguilla* L. following exposure to [beta]-naphthoflavone--a microsomal enzyme inducer. *Envir Interna* 2005;31(1):99-104.
- Bernier NJ, Peter RE. The hypothalamic-pituitary-interrenal axis and the control of food intake in teleost fish. *Compar Biochem Physiol Part B: Biochem Mol Biol* 2001;129(2): 639-44.
- Power DM, Llewellyn L, Faustino M, Nowell MA, Bjornsson BT, Einarsdottir IE, et al. Thyroid hormones in growth and development of fish. *Compar Biochem Physiol Part C: Toxicol Pharmacol* 2001;130(4):447-59.

9. Van Anholt RD, Spanings T, Koven W, Bonga SEW. Effects of acetylsalicylic acid treatment on thyroid hormones, prolactins, and the stress response of tilapia (*Oreochromis mossambicus*). *Am J Physio-Regu, Integ Compar Physio* 2003;285(5):1098-106.
10. Belanger J, Son JH, Laugero KD, Moberg GP, Doroshov SI, Lankford SE, Cech J. Effects of short-term management stress and ACTH injections on plasma cortisol levels in cultured white sturgeon, *Acipenser transmontanus*. *Aquacul* 2001;203(1):165-76.
11. Fontainhas-Fernandes A, Monteiro M, Gomez E, Reis-Henriques MA, Coimbra J. Effect of dietary sodium chloride acclimation on growth and plasma thyroid hormones in tilapia *Oreochromis niloticus* (L.) in relation to sex. *Aquacul Res* 2000;31(6):507-17.
12. Gad NS, Saad AS. Effect of environmental pollution by phenol on some physiological parameters of *Oreochromis niloticus*. *Global Vet* 2008;2:312-9.
13. Escartin E, Porte C. Assessment of PAH pollution in coastal areas from the NW Mediterranean through the analysis of fish bile. *Marine Pollu bulletin* 1999;38:1200-6.
14. Pacheco M, Santos MA. Induction of Liver EROD and Erythrocytic Nuclear Abnormalities by Cyclophosphamide and PAH in *Anguilla anguilla*L. *Ecotoxicol Envir Safety* 1998;40(1):71-6.
15. Wilson JM, Vijayan MM, Kennedy C, Iwama G, Moon TW. Beta-naphthoflavone abolishes interrenal sensitivity to ACTH stimulation in rainbow trout. *J Endocrin* 1998;157:63-70.
16. Barciela P, Soengas J, Rey P, Aldegunde M, Roza G. Carbohydrate metabolism in several tissues of rainbow trout, *Oncorhynchus mykiss*, is modified during ovarian recrudescence. *Compar Biochem Physio Part B: Compa Biochem* 1993;106(4):943-8.
17. Barry TP, Lapp AF, Kayes TB, Malison JA. Validation of a microtitre plate ELISA for measuring cortisol in fish and comparison of stress responses of rainbow trout (*Oncorhynchus mykiss*) and lake trout (*Salvelinus namaycush*). *Aquacul* 1993;117(3): 351-63.
18. Thomas P, Woodin B, Neff J. Biochemical responses of the striped mullet *Mugil cephalus* to oil exposure I. Acute responses—Interrenal activations and secondary stress responses. *Marine Bio* 1980;59(3):141-9.
19. Kennedy CJ, Farrell AP. Ion homeostasis and interrenal stress responses in juvenile Pacific herring, *Clupea pallasii*, exposed to the water-soluble fraction of crude oil. *J Exp Marine Bio Eco* 2005;323:43-56
20. Tintos A, Gesto M, Míguez JM, Soengas JL.  $\beta$ -Naphthoflavone and benzo (alpha) pyrene treatment affect liver intermediary metabolism and plasma cortisol levels in rainbow trout *Oncorhynchus mykiss*. *Ecotoxicol Environ Saf* 2008;69(2):180-6
21. Nguyen LP, Bradfield CA. The search for endogenous activators of the aryl hydrocarbon receptor. *Chem Res Toxicol* 2007;21:102-116.
22. Vijayan MM, Aluru N, Leatherland JF. 6 Stress Response and the Role of Cortisol. *Fish Dis Disorder* 2010;2:182.
23. Liu J, Liu Y, Barter RA, Klaassen CD. Alteration of thyroid homeostasis by UDP-glucuronosyltransferase inducers in rats: a dose-response study. *J Pharmacol Exp Therapeut* 1995; 273(2):977-85.
24. Hood A, Allen ML, Liu YP, Liu J, Klaassen CD. Induction of T4 UDP-GT activity, serum thyroid stimulating hormone, and thyroid follicular cell proliferation in mice treated with microsomal enzyme inducers. *Toxicol Appl Pharmacol* 2003;188(1):6-13.
25. Alkindi AYA, Brown JA, Waring CP, Collins JE. Endocrine, osmoregulatory, respiratory and haematological parameters in flounder exposed to the water soluble fraction of crude oil. *J Fish Biol* 1996;49(6):1291-305.
26. Stephens S, Alkindi A, Waring C, Brown J. Corticosteroid and thyroid responses of larval and juvenile turbot exposed to the water soluble fraction of crude oil. *J Fish Biol*1997;50(5): 953-64.
27. Waring C, Brown JA, Collins JE, Prunet P. Plasma prolactin, cortisol, and thyroid responses of the brown trout (*Salmo trutta*) exposed to lethal and sublethal aluminium in acidic soft waters. *Gen Compa Endocrin* 1996; 102:377-85.