Antimicrobial Activity of Five Medicinal Plants on *Candida Albicans*

Fatemeh Masomi, Mehdi Hassanshahian*

**ABSTRACT**

**Background:** In recent years, drug resistance to human pathogenic fungi has been increased. Medicinal plants are one way to overcome antibiotic resistance. The aim of this study was to evaluate the antifungal and inhibitory activity of five medicinal plants on the growth of *Candida albicans*.

**Methods:** This study was done in the Microbiology Lab of Shahid Bahonar University of Kerman, Iran in 2015. Five medicinal plants include: *Trachyspermum ammi* (seed), *Teucrium polium* (leaf), *Piper nigrum* (seed), *Pistachia vera* (skin), *Camelia sinensis* (leaf) were collected. Collected plant materials were extracted by ethanol and methanol solvent with maceration method. Antifungal activity of the ethanolic and methanolic extracts was evaluated by paper disc diffusion and agar well diffusion methods. Besides, MIC and MBC of each extract was determined.

**Results:** All plant extracts had sufficient inhibitory effect against *C. albicans* but the extracts of *P. vera* had the best inhibitory effect on *C. albicans* (ZOI: 40 mm). The lowest antifungal effect between these five plants related to *Piper nigrum* (ZOI: 13 mm). Besides, the *P. vera* extracts had the best MIC and MBC values (6.25 and 12.5 mg/ml).

**Conclusion:** This study strongly evidence the maximum antimicrobial activity of medicinal plants against *C. albicans* that this inhibitory effect varies with the different solvent-extract form. A more comprehensive study need to identify the effective compounds that have these antifungal properties.

**Keywords:** Antibiotic, Antifungal Activity, Extracts, Medicinal Plants, Inhibitory Effect.

**INTRODUCTION**

Despite extensive progress in scientific knowledge and medical technology, infectious diseases remain a leading cause of worldwide morbidity and mortality. The main chemical agents for eradication of pathogenic fungal and bacteria are antibiotics. For many times antibiotics was gold key for treatment of pathogens. However, in recent years antibiotic resistant microbes was increased and the use of antibiotics for treatment of pathogenic microbes had less efficiency [1]. *Candida albicans* is an oral commensal flora that causes opportunistic local and systemic infections in immune compromised individuals [2]. Due to the development of resistance in known fungal pathogens and the emergence of fungal pathogens intrinsically resistant to the currently available antibiotics, it is important that novel antifungal agents be identified and developed [34]. Natural products are sources of molecules that can be used as antimicrobial agents, an attempt to overcome drug resistance to old and new antimicrobials used currently in clinical therapy [5]. In addition, it is expected that plant compounds showing target sites other than those currently used by antibiotics will be active against drug-resistant microbial pathogens [6].

*Trachyspermum ammi* (L.) Sprague (TA) from the family Apiaceae is an old herb with various medical properties. There are medieval and traditional reports on the effects of *T. ammi* oil and hydrosol on neural disorders such paralysis, tremor, and palsy as well as chronicpains. *T. ammi* was also reported as an aphrodisiac, galactagogue, and diuretic agent. Persian practitioners also reported the use of the fumigation form of *T. ammi* seeds in female genital disorders [7].

The genus *Teucrium*, which belongs to the family Labiatae, includes 300 species widespread all around the world [8, 9]. A large number of known medicinal species belonging to the genus *Teucrium* are used in folk medicine and pharmacy [10].

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Black pepper (*Piper nigrum* L.) is a flowering vine of the Piperaceae family cultivated for its fruit, usually dried and used as a spice and seasoning. Pepper is also used in folk medicine as aphrodisiac, carminative, stomachic, antiseptic diuretic and for the treatment of cough, rheumatoid arthritis, peripheral acetic acid, neuropathy, melanoderma and leprosy due to the presence of volatile compounds, tannins, phenols and other unknown substances [11].

Among the aromatic plants belonging to the family of Anacardiaceae, the genus *Pistacia* is noteworthy for its numerous species and varieties of wild-growing plants. Many of these species are typical of the Mediterranean area. *Pistacia* has an economic value as it is the source of traditional medicinal agent "gum" mastic, an oleoresin exudate from the stem of this plant [12]. It is a traditional natural remedy that has been used by very ancient Mediterranean civilizations like Greeks and Egyptians [13].

Tea is a very popular drink world-wide. It is produced from the plant *Camellia sinensis*, grown in at least 30 countries, and grows best in certain tropical and subtropical regions [14]. Most of the black tea produced is consumed in the United States, Oolong tea is most popular in China and Taiwan, and green tea is most popular in China, Japan, and Korea [15]. Among the health benefits studied using green tea are: as an antioxidant, anti-inflammatory, anticarcinogenic, in cardiovascular health, oral health, and as an antimicrobial. Antioxidant effects come from the ability of green tea to limit the amount of free radicals by binding to reactive oxygen species (ROS) [16].

The aim of this study was evaluation the antifungal and inhibitory activity of these five mentioned medicinal plants on the growth of *C. albicans*.

**MATERIAL AND METHODS**

**Plant Materials**

The five plants, namely *T. ammi* (seed), *T. polium* (leaf), *P. nigrum* (seed), *P. vera* (skin), *C. sinensis* (leaf) were collected. The taxonomical identification of the plant was confirmed by Dr. Mirtajadini at the Department of Biological Sciences, Shahid Bahonar University of Kerman, Kerman, Iran.

**Preparation of Plant Extracts**

Collected plant materials were air dried and pulverized into fine powder. Fifty gr of each powder of plants was macerated in 500 ml of ethanol and methanol solvents. Different extracts were prepared using the cold maceration process for 72 h at room temperature under constant shaking and filtered with Whatman No.1 filter paper. The residue was further macerated twice with the same solvent overnight and filtered. The filtrates obtained from each extraction were mixed and concentrated under vacuum. The extracts obtained were kept at 4°C for further use [8, 11].

**Determination of Antibacterial Activity**

**Disk Diffusion Method**

Antifungal activity of the ethanolic and methanolic extracts was evaluated by paper disc diffusion method. Stock culture of test fungus was grown in PDB (Potato Dextrose Broth) medium at 37 °C for 24 h. Final cell concentrations were adjusted to 10^5 cfu/mL with reference to the McFarland turbidimeter. One mL of this inoculum was added on the surface of each plate containing Mueller-Hinton agar (MHA, Oxoid) by sterile cotton swab and allowed to remain in contact for 1 min then, 0.15 mg/ml concentration of each extract prepared from sterile 6mm filter paper discs were placed into each of these concentrations for 1 h. The disc put for 30 minutes at room temperature and transferred to the medium. Disc solvent-free extract used as positive control. The inhibition zone around each disc was measured in millimeter and the assay was carried out three times for each extract [12, 15].

**Agar Well Diffusion Method**

Agar well-diffusion method was followed to determine the antifungal activity. Potato Dextrose Agar (PDA) plates were swabbed (sterile cotton swabs) with 8 h old-broth culture of *C. albicans*. Wells (10 mm diameter and about 2 cm a part) were made in each of these plates using sterile cork borer. Stock solution of each plant extract was prepared at a concentration of 1 mg/ml in different plant extracts viz. methanol and ethanol. About 40 µl of different concentrations of plant solvent extracts were added sterile syringe into the wells and allowed to diffuse at room temperature for 2 h. Control experiments comprising inoculums without plant extract were set up. The plates were incubated at 34 °C for 24 h. The diameter of the inhibition zone (mm) was measured and the activity index was also calculated. Triplicates were maintained and the experiment was repeated thrice, for each replicates.
the readings were taken in three different fixed directions and the average values were recorded [5, 9].

**Determination of Minimal Inhibitory Concentration (MIC), and Minimal Fungicidal Concentration (MBC)**

The minimum inhibitory concentrations (MIC) were performed by a serial dilution technique using 96-well microtiter plates. The different plant extracts viz. Methanol and Ethanol were taken (1 mg/ml) and serial dilution of the extract with potato dextrose broth medium with respective inoculum were used. The microplates were incubated for 24 h at 34 °C, respectively. The lowest concentrations without visible growth (at the binocular microscope) were defined as MICs. The MBC was determined by spreading 50 µl on PDA plate from the sample showing no visible growth and it was further incubated for 18 h at 37 °C [9, 11].

**RESULTS**

**Evaluation of the Antifungal Activities of Methanolic and Ethanolic Plant Extracts against C. albicans**

The zone of inhibition (ZOI) for methanolic and ethanolic extracts of five tested plants that assayed by disc diffusion method are shown in Table 1, also the results of agar well diffusion method of these extracts are illustrated in Table 2. All plant extracts had sufficient inhibitory effect against C. albicans but the extracts of P. vera have the best inhibitory effect on C. albicans (ZOI: 40 mm). The methanolic extracts of these five herbal plants were better than ethanolic extracts. The lowest antifungal effect between these five plants related to P. nigrum (ZOI: 13 mm). The ZOI in agar well diffusion was higher than disc diffusion method in all tested plants.

**MIC and MBC Values of Five Plant Extracts against C. albicans**

The results of MIC and MBC values of the five plant extracts were illustrated in Table 3. Approximately all plant extracts exhibited inhibitory effect in lower concentration. However, the P. vera extracts had the best MIC and MBC values (6.25 and 12.5 mg/ml). Some plant methanolic extracts such as T. ammi and C. sinensis had not MIC value and only exhibit MBC value. It is means that these plants extracts only kill C. albicans and not inhibited this fungus. Considering that these extracts used in broth media in MIC test, in the lower concentration that used in preparing disks (0.156-2.5 mg/ml) and inhibited C. albicans, the inhibitory efficiency of these extracts in broth medium is more than solid medium.

<table>
<thead>
<tr>
<th>Type of Plant</th>
<th>Candida albicans (ZOI in mm)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Ethanol Extract</td>
</tr>
<tr>
<td><em>Trachyspermum ammi</em></td>
<td>13 ± 1.1</td>
</tr>
<tr>
<td><em>Teucrium polium</em></td>
<td>11 ± 0.9</td>
</tr>
<tr>
<td><em>Piper nigrum</em></td>
<td>11 ± 0.6</td>
</tr>
<tr>
<td><em>Pistachia vera</em></td>
<td>17 ± 0.7</td>
</tr>
<tr>
<td><em>Camelia sinensis</em></td>
<td>14 ± 1.3</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Type of Plant</th>
<th>Candida albicans</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>MIC (mg/ml)</td>
</tr>
<tr>
<td><em>Trachyspermum ammi</em></td>
<td>12.5</td>
</tr>
<tr>
<td><em>Teucrium polium</em></td>
<td>12.5</td>
</tr>
<tr>
<td><em>Piper nigrum</em></td>
<td>25</td>
</tr>
<tr>
<td><em>Pistachia vera</em></td>
<td>6.25</td>
</tr>
<tr>
<td><em>Camelia sinensis</em></td>
<td>12.5</td>
</tr>
</tbody>
</table>

Table 1. The inhibitory effect of five medicinal plants extracts against C. albicans that assayed by disc diffusion method.

Table 2. The anti-Candida effects of five medicinal plants extract that assayed by agar well diffusion method.
The results of this study showed the best antimicrobial activity of medicinal plants. These results corroborate the importance of ethno-pharmacological surveys in the selection of plants for bioactivity screening. The results obtained represent a worthwhile expressive contribution to the characterization of the anti-Candida activity of essential oils and plant extracts of traditional medicinal plants from the Iranian flora. Subsequently, bio-guided fractionation will be conducted on plants showing potential anti-Candida activity to identify the active compounds. Evaluations of the essential oils against other important human pathogens are also being conducted.

**CONCLUSION**

This study strongly evidences the maximum antimicrobial activity of medicinal plants against *C. albicans* that this inhibitory effect varies with the different solvent-extract form. A more comprehensive study need to identify the effective compounds that have these antifungal properties.

**ACKNOWLEDGMENTS**

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**REFERENCES**


