Protective Effect of *Nigella Sativa* (Black Caraway) Oil on Oral Dichlorvos Induced Hematological, Renal and Nonspecific Immune System Toxicity in Wistar Rats

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ABSTRACT

**Background:** Exposure to environmental toxins such as organophosphates poses a great threat to the health of the public. In this work, we investigated the effects of continuous exposure to dichlorvos (DDVP) on kidney function and hematological parameters, and the possible antidote activity of *Nigella sativa* oil (NSO).

**Methods:** This research was conducted in 2016, at The Animal Holding and Research Laboratory of Faculty Basic Medical Sciences, University of Ilorin, Ilorin, Nigeria. Twenty-four Wistar rats were randomly divided into four groups, six rats each. The four groups received: 1. phosphate buffer solution as controls, 2. DDVP, 3. DDVP+NSO and 4. NSO alone. After 2 wk of treatment, blood samples were collected and hematological profile (RBC, Hb), erythrocyte indices (MCV, MCH, MCHC, and Plt), renal function parameters (albumin, urea, total protein, chloride, sodium, and potassium ions) and nonspecific immune response (WBC) were measured.

**Results:** Rat exposed to DDVP showed red blood cell count, hemoglobin, packed cell volume, albumin, and total protein levels was reduced from control, while white blood cell count and urea significantly increased as compared to controls, the change in K$^+$ level was not significant. NSO maintained optimal levels of red blood cell count, hemoglobin, packed cell volume, albumin, white blood cell count, and urea, indicative of its protective effect against hemato- and nephrotoxicity of DDVP.

**Conclusion:** *N. sativa* (Black Caraway) oil might be a potential antidote in hematoxicity, immunosuppression and renal dysfunction in organophosphate poisoning, especially dichlorvos. The protective effect of NSO against dichlorvos toxicity can be attributed to its antioxidant capacity.

**Keywords:** Anemia, Dichlorvos, Kidney Function Tests, *Nigella Sativa*, Organophosphates, Organophosphate Poisoning.

INTRODUCTION

Harmful chemicals in the environment and their burden on health have become an issue of great importance in recent decades [1], causing toxicity to a variety of biota and human life [2].

Organophosphates (OPs) are one of the most dangerous chemicals used in agriculture and household, and are highly toxic to liver and muscles, as well as, nervous, immune, urinary, reproductive and hematological systems [3-5].

A commonly used OP, dichlorvos or 2, 2-dichlorovinyl dimethyl phosphate (DDVP), is extensively used for pest and insect control [6], and despite the reported toxicity, it is a major constituent of most of the insecticides used in the developing world [7]. Although several antidotes are available in the management of OP toxicity, the available options have some limitations with unavoidable side effects [8]. Therefore, the search for new and novel therapeutic drugs for OP poisoning is crucial. Alternative medicine and supplementary substances with beneficial effects in...
immunological, hematological and renal systems definitely have a place in the treatment of OP poisoning.

In recent years, interest in the use of natural products as alternative therapy in disease conditions has increased, gaining wide acceptance from the public and medical professionals. *Nigella sativa Linn.* (Ranunculaceae) is a medicinal plant also known as black caraway, black cumin, *Habbatal Barakah* or and Kalonji seed, is widely used in the treatment of various ailments such as bronchial asthma, cough, diarrhoea, abdominal pain, and dyslipidemia [9]. *N. sativa* oil (NSO) exhibits pharmacological activities including the ability to act as a chelating agent [10], antioxidant and anti-inflammatory [11], immunomodulatory and antitumor substances [12].

Blood, being the medium of intercellular transport, plays an important role in the immune system. It rapidly comes in direct contact with various tissues in the body, therefore, the physiological state of an organism is always reflected in the contents of the blood [13], and thus, the hematological and biochemical parameters can indicate toxicity with great potential for environmental monitoring of health [14]. Perturbations in immune functions are caused by immunotoxic compounds, leading to immune suppression, resulting in decreased resistance to infections, hypersensitivity, and autoimmunity caused by disorders in immune regulation [13].

This study investigated the antidotal efficacy of *N. sativa* oil in mitigating dichlorvos induced hematological, immunological and renal damage in Wistar rats.

**MATERIALS AND METHODS**

**Chemicals and Drugs**

Dichlorvos was purchased from the Sigma Chemicals (St. Louis, MO, USA), while the phosphate buffer solution (PBS) was prepared in our laboratory. The black caraway oil (100% pure natural oil) was obtained from Masrawarda, Kingdom of Saudi Arabia.

**Animals**

Twenty-four adult male Wistar rats with an average weight of 200 ± 20 gr were used. The animals were housed (six per cage) under standard laboratory conditions in the animal holding of the Faculty of Basic Medical Sciences, University of Ilorin, Nigeria, in 2016. They were allowed free access to water and food *ad libitum.*

**Treatments Schedule**

The rats were randomly divided into five groups (n = 6) as follows:

- **Controls:** received phosphate buffer solution (PBS) (1 ml/kg oral)
- **Exp. 1:** received DDVP (8.8 mg/kg/day orally) [15]
- **Exp. 2:** received DDVP (8.8 mg/kg/day orally) + NSO (1 ml/kg oral) 30 min. post-treatment
- **Exp. 3:** received NSO (1 ml/kg orally) [16]

All administrations were scheduled and carried out during the light phase between 7:00 and 9:00 am. All groups contained six rats each and treatments continued for fourteen consecutive days.

**Ethical Approval**

All experimental procedures were performed in accordance with our institutional guidelines for animal care and use and ethical approvals were received from the University of Ilorin's Ethics Committee.

**Hematological, Immunological and Erythrocytes Indices Study**

Animals were euthanized with intraperitoneal ketamine injection, and after immobility was confirmed, blood samples were collected. Freshly collected blood samples were analyzed for hematological analysis using an automatic hematological assay analyzer (Beckman Coulter, USA). Hematological parameters that were tested were as follows: red blood cells count (RBC), packed cell volume (PCV), white blood cells count (WBC), mean cell volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), hemoglobin (Hb), neutrophils and lymphocytes counts.

**Kidney Function Study**

Blood samples were centrifuged at 3000 rpm for 15 min. The plasma was collected and used for the analysis of urea, albumin, total protein, Na⁺, K⁺, and Cl⁻ using commercial kits supplied by Randox Laboratories Limited, United Kingdom.

**Statistical Analysis**

Data recorded in this study were reported as mean ± standard error of mean. They were analyzed using one-way analysis of variance (ANOVA) and for post-hoc analyses, we used the Bonferroni test. A *P*-value of ≤ 0.05 was considered statistically significant.
RESULTS

Effect of DDVP and NSO on Hematological Parameters

Dichlorvos (DDVP) caused a significant (P≤0.05) reduction in red blood cells count (RBC), hemoglobin (HGB) and packed cell volume (PCV), while post-treatment with NSO increased all these indices (Table 1).

Effects of DDVP and NSO on Erythrocytes Indices

DDVP resulted in reduction in the mean cell volume (MCV), mean cell hemoglobin (MCH), and increased mean cell hemoglobin concentration (MCHC), although not statistically significant. While NSO only and DDVP rats that were post-treated with NSO demonstrated fair levels of MCV, MCH and MCHC (Table 1).

Table 1. Showing levels of hematological parameters, Erythrocytes indices and Peripheral blood leucocytes in animals treated with PBS, DDVP, DDVP+NSO and NSO only.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Hematological Parameters (x10¹²/L)</th>
<th>Erythrocytes Indices (fl)</th>
<th>Peripheral Blood Leucocytes (X10⁹/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RBC</td>
<td>HGB</td>
<td>PCV (%)</td>
</tr>
<tr>
<td>PBS</td>
<td>6.74±0.13</td>
<td>16.08±0.7</td>
<td>42.20±1.4</td>
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<td></td>
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<tr>
<td>DDVP</td>
<td>3.54±0.24*</td>
<td>7.86±0.52</td>
<td>23.80±2.0</td>
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<tr>
<td>DDVP+NSO</td>
<td>6.19±0.25</td>
<td>15.62±0.4</td>
<td>41.00±1.4</td>
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<tr>
<td>NSO</td>
<td>6.81±0.14</td>
<td>14.06±0.9</td>
<td>42.60±1.2</td>
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<tr>
<td>ANOVA*</td>
<td>= P≤0.05</td>
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Table 2. Showing plasma levels of kidney function parameters in rats treated with PBS, DDVP, DDVP+NSO and NSO only.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Alb (g/dl)</th>
<th>Urea (mmol/l)</th>
<th>Total Prot. (g/dl)</th>
<th>Na⁺ (mg/dl)</th>
<th>K⁺ (mg/dl)</th>
<th>Cl⁻ (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PBS</td>
<td>1.27±0.06</td>
<td>11.68±0.34</td>
<td>5.57±0.47</td>
<td>29.64±4.33</td>
<td>3.94±0.49</td>
<td>67.15±15.67</td>
</tr>
<tr>
<td>DDVP</td>
<td>0.75±0.63*</td>
<td>16.33±0.38⁷</td>
<td>5.40±0.59</td>
<td>34.34±5.69</td>
<td>8.76±1.61*</td>
<td>69.54±2.37</td>
</tr>
<tr>
<td>DDVP+NSO</td>
<td>1.22±0.08</td>
<td>13.58±0.61</td>
<td>5.48±0.34</td>
<td>30.64±6.04</td>
<td>8.40±1.18</td>
<td>62.42±5.98</td>
</tr>
<tr>
<td>NSO</td>
<td>1.12±0.07</td>
<td>9.79±0.50⁷</td>
<td>6.08±0.26</td>
<td>23.12±2.78</td>
<td>6.64±0.43</td>
<td>55.63±6.14*</td>
</tr>
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</table>

ANOVA* = P≤0.05

DISCUSSION

Indiscriminate and uncontrolled use of pesticides and insecticides have resulted in severe environmental pollution, acute and chronic human poisoning. The bioaccumulation of these substances is associated with metabolic, immunological, oxidative and antioxidative changes, and can reduce human lifespan [17].

Blood is a pathological and physiological indicator [18] and chemicals, enzymes or ion concentrations in the blood or its derivatives are validated markers of health. The reduced levels of RBC, HGB, and PCV reported in this study, following DDVP exposure, suggested the hematotoxic effects of DDVP. This was in accordance with a previous study [19] and was strengthened by the reports concerning exposure to other OPs [13, 20, 21].

Red blood cell indices reflect the size (MCV) and hemoglobin content (MCH and MCHC) of red
blood cells and aid in the diagnosis of possible anemia inducing properties of different substances. The reduction in MCV and MCH with an increase in MCHC that we observed, were similar to previous researches [13, 19], but it was in disagreement with the report that claimed an undisturbed erythrocytes indices after exposure to OP [22]. Also, the changes in total WBC, neutrophils, and lymphocytes documented in this study were in where strengthened with the reports of previous studies [13, 14], with the exception of a reduction in lymphocyte count [13].

In the present study, levels of serum albumin, urea, total protein, chloride, sodium and potassium ions were also measured. Some impaired levels of these parameters following DDVP exposure suggested renal dysfunctions in the treated animals, and this was similar to what has been extensively reported in the scientific literature following exposures to most insecticides [3, 4, 23]. The elevation of urea concentrations may be attributed to a reduction in the glomerular filtration rate in the kidney [24], and the reduction in the albumin and total protein levels are significant indications of renal dysfunctions, as reported recently with OPs [13, 25].

Post-treatment with NSO neutralized DDVP associated hematological, immunological and renal toxicity. These effects can be attributed to NSO's efficacy against OP-induced toxicity in various body systems including hormonal, reproductive, liver and kidneys [26-29].

The kidney is the major target organ for exogenous toxicants, and it is important that any substance with possible therapeutic use against DDVP poisoning should protect renal function too. As observed in this study, NSO reduced serum urea concentration, thereby normalizing impaired urine filtration induced by DDVP. This finding was supported by a recent study employing NSO as a protective agent against cisplatin-induced nephrotoxicity in male rats [30].

The protective effect of N. sativa oil against dichlorvos toxicity can be attributed to its antioxidant capacity, as previous works have reported the beneficial effects of antioxidants drugs and supplements against OPs poisonings, including DDVP [31-33].

CONCLUSION

N. sativa (Black caraway) oil might be a potential antidote in hematotoxicity, immunosuppression and renal dysfunction in organophosphate poisoning, especially dichlorvos.

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REFERENCES

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