Reduction of Aflatoxin M1 in Milk Using Kefir Starter

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ABSTRACT

Background: Mycotoxins naturally occur in foods. Aflatoxins can cause serious health problems in consumers. Nowadays, biological detoxification method is considered to decrease the aflatoxins level in foods. The aim of this study was to evaluate the effect of kefir starter microorganisms to decrease the aflatoxin M1 (AFM1) levels in milk.

Methods: The study was carried out at Shabestar branch, Islamic Azad University in 2016. AFM1 at three levels 150, 200 and 250 ng/L was added to milk samples. Then a pool of lactic acid bacteria (LAB), yeasts and full kefir starter culture was added to milk samples. After cool storage of samples in 4 °C for 7 d, all samples were collected and the level of AFM1 determined by HPLC method. All samples were prepared in triplicate.

Results: The highest reduction percentage of AFM1 was observed in yeast (65.33%-68.89%) and LAB pool (65%). Samples with full kefir starter showed the reduction percent range of 11.67-34.66% that was lower in compare with other treatment groups.

Conclusion: These findings support the ability of LAB and yeasts to bind to aflatoxins in foods. Kefir drink in countries with high contamination by AFM1 in milk can be a safe dairy product choice for consumers.

Keywords: Aflatoxin M1, Detoxification, Kefir, Lactic Acid Bacteria, Yeast.
irradiation, microbial inactivation, and fermentation have been suggested by researchers. Some strains of lactic acid bacteria have the ability to bind aflatoxins in foods [10]. The mechanism of action is not clear. An adhesion to bacterial is the effect of lactic acid bacteria (LAB) on the aflatoxin reduction in fermented products. Some researchers have studied the ability of bacteria to decrease aflatoxins in milk and dairy products [10-12].

The aim of this research was to evaluate the kefir starter effects on the level of AFM1 in cow milk during kefir drink production and storage of kefir drink at 4 °C.

**MATERIALS AND METHODS**

**Chemicals**

The study was carried out at Shabestar branch, Islamic Azad University in 2016. Aflatoxin M1 was prepared from Sigma (Germany). AFM1 levels were 150, 200 and 250 ng/L. Each level was repeated for three times.

**Kefir Starter Microorganisms**

Kefir starter was prepared from CHR Hansen-Denmark. Three packages included: 1) F-DVS ABT-2 containing *Lactobacillus acidophilus,* *Bifidobacterium* BB-12, and *Streptococcus thermophilus* (thermophilic lactic culture); 2) FD-DVS CHN-22 containing *Lactococcus lactis subsp.* *Cremoris,* *Leuconostoc,* *Lactococcus lactis subsp.* *Lactis biovar diacetylactis,* *Lactococcus lactis subsp.* *Lactis*; 3) LAF containing *Debaryomyces hansenii,* *Kluyveromyces marxianus subsp.* *Marxianus*.

For packages 1 and 2 of starter’s packages, the incubation temperature was 22 °C for 6 h. Then the yeasts package was added and incubated at room temperature for production of gas in kefir drink.

**Kefir Drink Preparation**

Kefir drink production was according to the CHR Hansen procedure. For study, each package and full kefir starter were compared.

Kefir starter was added to cow milk (Nestle-Iran) that was free of AFM1. One hundred grams of milk powder were added to 1000 ml of deionized water, and milk was heated at 120 °C for 15 min and 0.01 gr of each starter’s packages were added to 1000 mL of milk.

AFM1 was added at three levels to milk and kefir starter microorganisms were added to three types (Treatment) included *Lactobacilli* + *Lactococci* (T1), yeasts (T2) and full kefir starter (T3). For each level of AFM1 control sample(C) was prepared. Kefir starter microorganisms were not added to control samples. The milk was allowed to ferment and the kefir fermentation was considered complete with pH of 4.6. Samples were collected after 7 d maintenance at 4 °C. The amount of AFM1 was detected by HPLC method. All samples were prepared in triplicate. Samples’ pH determination was carried out according to ISIRI. No.2852 with pH meter (HORIBA, Japan) at 1th and 7th d.

**HPLC Method**

The HPLC method was validated according to ISIRI. No. 7133. Three steps included extraction, purification, and quantitative determination was used to measure the AFM1 level in samples by HPLC. Samples fatty layer was removed by centrifugation at 2500 gr for 10 min. Waters HPLC system (Waters co., Milford, MA. USA) with Waters 2475 detector were used for AFM1 detection. The analytical column was C18 column (ODS-2) [13,14].

**Statistical Analysis**

For statistical analysis SPSS software ver. 16 (Inc., Chicago, IL, USA) was used and ANOVA and independent *t*-test were performed. Results with *P*<0.05 were considered statistically significant.

**RESULTS**

Comparing three treatment groups of microorganisms with statistical analysis showed that there were significant differences between the amounts of AFM1 in all strains. The results in Table 1 show the effect of pH on AFM1 reduction in samples treated by microorganisms. The overall rates for AFM1 in kefir drink have been shown in Table1.

The reduction rate of AFM1 kefir treatments has been shown in Figure 1. The highest reduction percentage was seen in LAB pool and yeast strains with the different amount of initial AFM1. The initial amount of AFM1 at the common level of the strains’ amount in milk is not effective on the degradation percentage of AFM1 in samples.

The full stains’ samples compared to each strain group showed the lower reduction percentage of AFM1 (Figure 1).
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Table 1. Average mean values of AFM1 in kefir treatments, T1: Lactobacilli + Lactococci, T2: yeasts, T3: full kefir starter.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Initial AFM1 in milk(ng/L)</th>
<th>pH</th>
<th>Average Mean values of AFM1 in kefir treatments(ng/L) mean±SEM</th>
<th>pH</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>150</td>
<td>5.84</td>
<td>150</td>
<td>5.94</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>6.07</td>
<td>200</td>
<td>6.05</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>250</td>
<td>6.05</td>
<td>250</td>
<td>4.93</td>
<td>-</td>
</tr>
<tr>
<td>T1</td>
<td>150</td>
<td>4</td>
<td>63.33±3.33**</td>
<td>3.94</td>
<td>0.001</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>4.04</td>
<td>70±5.77**</td>
<td>4.04</td>
<td>0.001</td>
</tr>
<tr>
<td></td>
<td>250</td>
<td>4.07</td>
<td>113.33±13.33</td>
<td>3.95</td>
<td>0.009</td>
</tr>
<tr>
<td>T2</td>
<td>150</td>
<td>4.17</td>
<td>46.66±8.81</td>
<td>4.18</td>
<td>0.007</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>4.21</td>
<td>110±15.27</td>
<td>4.52</td>
<td>0.028</td>
</tr>
<tr>
<td></td>
<td>250</td>
<td>4.38</td>
<td>86.66±12.01*</td>
<td>4.68</td>
<td>0.005</td>
</tr>
<tr>
<td>T3</td>
<td>150</td>
<td>4.02</td>
<td>120±11.54</td>
<td>3.98</td>
<td>0.060</td>
</tr>
<tr>
<td></td>
<td>200</td>
<td>4.10</td>
<td>176.66±3.33*</td>
<td>4.01</td>
<td>0.020</td>
</tr>
<tr>
<td></td>
<td>250</td>
<td>4.02</td>
<td>163.33±6.66*</td>
<td>3.97</td>
<td>0.006</td>
</tr>
</tbody>
</table>

**P<0.001, *P<0.05

DISCUSSION

The percentages of AFM1 reduction in kefir drink treatments in the present study are in agreement with the results of others [10,15]. The maximum reduction rate of AFM1 was at 20% kefir sample and between bacterial samples; L. casei had the maximum reduction. The added yeast to LAB was increased the reduction rate of AFM1 in samples [16]. At present study, the maximum reduction was in samples with yeast. Samples contained LAB (lactobacilli + lactococci) showed high reduction rate of AFM1 at the toxin group involved of 200 ng/L.

The ability of S. cerevisiae strain was studied and a pool of lactic acid bacteria strains in reduction of AFM1 in UHT skimmed milk. Using S. cerevisiae with LAB pool increased the level of AFM1 during 60 min by 100% [15]. In our study, Debaryomyces hansenii and Kluyveromyces marxianus subsp. Marxianus, showed the highest reduction of AFM1 level. Yeast strains have been investigated as an aflatoxin binder[17]. Studies mainly focus on S. cerevisiae. The reduction rate depends on strain specific and varies among different strains[17].

Table 2 gives a range of reduction percentage in aflatoxins level from some reports about the effects of fermentation on aflatoxins in milk.

The reduction of AFM1 in different kefir drink might be due to binding AFM1 to bacterial cell wall components and low pH in kefir [18]. Low pH can cause to degradation of AFM1 in kefir drink. On the other hand, the oxidation of glucose produces hydrogen peroxide in kefir drink and the reactive oxygen will react with AFM1 molecule[10].
The effect of pH on in vitro reduction of aflatoxin B1 was studied by strains of LAB isolated from Moroccan sourdough bread and the highest removal of AFB1 was at pH range from 3 to 4.5 [19]. Yogurt fermented by 50% yogurt culture and 50% L. planetarium had the highest reduction at the rate of AFM1 at the end of a storage period. Similar results was obtained at present study [10].

Different strains of LAB had distinct reduction percentage of AFM1. There are some reports that the fermentation showed no effect at the level of AFM1 in foods [18]. Our findings support the ability of LAB and yeasts to bind to mycotoxins in foods.

Table 2. Examples of reported effects of microorganism's fermentation on aflatoxins.

<table>
<thead>
<tr>
<th>Toxin type</th>
<th>Raw materials</th>
<th>Product</th>
<th>Type of fermentation</th>
<th>Percent of reduction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>Yogurt</td>
<td>lactic</td>
<td>69.8-87.8%</td>
</tr>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>Yogurt</td>
<td>lactic</td>
<td>26.2-34%</td>
</tr>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>UHT milk</td>
<td>lactic</td>
<td>11.7%</td>
</tr>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>UHT milk</td>
<td>Lactic+Yeast</td>
<td>100%</td>
</tr>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>UHT milk</td>
<td>Yeast</td>
<td>92.7%</td>
</tr>
<tr>
<td>Aflatoxin M1</td>
<td>milk</td>
<td>Kefir</td>
<td>Lactic+Yeast</td>
<td>91.91%</td>
</tr>
<tr>
<td>Aflatoxin B1</td>
<td>PBS</td>
<td>PBS</td>
<td>Lactic</td>
<td>5.6-25.7%</td>
</tr>
<tr>
<td>Aflatoxin B1</td>
<td>PBS</td>
<td>PBS</td>
<td>Lactic</td>
<td>50%</td>
</tr>
</tbody>
</table>

CONCLUSION

LAB and yeasts can be used in dairy product to reduce the AFM1 in milk. Furthermore, the kefir drink use can improve the health conditions of dairy consumers. The mechanism of removal effect of yeast is not clear and additional studies are needed. Overall, microorganisms in the kefir starter have the ability to reduce the AFM1 amount to safe level in countries that the milk and dairy products have shown high contamination by AFM1 like as Iran. Although the kefir starter reduces toxicity, it is not able to complete detoxification. Additional studies are recommended to investigate the other strains of bacteria and yeasts’ mechanism on mycotoxins reduction in foods.

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The authors declare that there is no conflict of interest.

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