Research Paper:
Effect of Sodium Benzoate on Apoptosis and Mitochondrial Membrane Potential After Aluminum Toxicity in PC-12 Cell Line

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Background: Sodium benzoate, a food preservative, prevents the growth of fungi and bacteria. Numerous studies have reported the protective effects of sodium benzoate on the nervous system. This study investigated the effect of sodium benzoate on cell apoptosis and mitochondrial function in an aluminum cell toxicity model.

Methods: After 48 hr of treating PC-12 cells with varying concentrations of sodium benzoate (0.125, 0.25 or 0.5 mg/ml) in the presence of aluminum maltolate (500 μM), the cell viability was assessed by MTT assay. The type of cell death (necrosis or apoptosis) was analyzed by flow cytometry (7-ADD/An V-PE staining). Also, rhodamine 123 was used to measure the Mitochondrial Membrane Potential (MMP). The acetylcholinesterase activity (AChE) was assessed by Ellman’s method.

Results: It was observed that sodium benzoate inhibited aluminum induced cell death within 48hr. Sodium benzoate at a concentration of 0.5 mg/ml significantly reduced the apoptotic cells that had been exposed to aluminum. Exposure of PC-12 cells with sodium benzoate and aluminum, increased the AChE activity, although, no significant changes in mitochondrial membrane potential were observed.

Conclusion: Sodium benzoate may provide its protective effects through increased AChE activity and inhibiting apoptosis induced by aluminum toxicity. It is likely that the disruption of MMP is neither involved in the induction of apoptosis by aluminum nor in the protective effect of sodium benzoate.

Keywords: Sodium benzoate, Aluminum, Neurotoxicity, PC-12 cell line, Necrosis and apoptosis, Acetylcholinesterase

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Introduction

The origin and etiologic of neurodegenerative disorders, such as Alzheimer’s and Parkinson’s Diseases (AD, PD) is multifactorial. Some basic metals are believed to contribute to the development of these disorders. One of the metals that is increasing known to play a major role in cell toxicity in the central nervous system is aluminum [1]. This metal may enter the human body via food, drinking water, beverages and medications [2, 3]. Some studies have shown that aluminum induces neuronal death and produces neurofibrillary tangles, as documented by AD studies [4, 5].
Mitochondria, a vital organelle in eukaryotic cells, have various roles including cellular energy generation, oxygen consumption, fatty acid metabolism, ions transport and storage, among others. Aluminum toxicity causes mitochondrial dysfunction, such as defective electron transport chain complexes [6, 7]. There is evidence to suggest that beta amyloids can disrupt mitochondrial functions directly by inducing neuronal death [8, 9]. Mitochondrial membrane potential (MMP) is reduced due to abundant presence of Reactive Oxygen Species (ROS), secondary to aluminum toxicity in neurons [10]. Further, it is known that the number of mitochondria declines during the development of AD [11].

Sodium benzoate is a food additive that is largely used due to its bacteriostatic and fungistatic properties under acidic condition [12]. It’s use is approved by the U.S. Food and Drug Administration, and there is evidence to show that it is useful to the management of diseases, such as urea cycle disorder in children [13], cognitive impairment in the early phase of AD and schizophrenic symptoms by inhibiting D-amino acid oxidase [14]. In addition, the protective effects of sodium benzoate have been shown in other diseases, such as multiple myeloma, multiple sclerosis and PD [15].

Acetylcholinesterase (AChE) is an important enzyme that serves in the normal function of cholinergic neurons in the central nervous system [16]. The cholinergic hypothesis has been proposed for the histopathological and biochemical variations of neurotransmitter markers in the brains of patients with AD [17]. The degeneration of cholinergic neurons has been observed in AD and inhibition of AChE activity is known to be the target in the treatment of this disease [18].

Previously, we have showed that sodium benzoate inhibits aluminum toxicity in PC-12 cell line [19]. This study was conducted to explore the role of sodium benzoate in apoptosis, and its impact on MMP and acetylcholinesterase activity cell toxicity with aluminum, as an experimental model of neurodegenerative diseases, such as AD.

Materials and Methods

Reagents & chemicals: PC-12 rat pheochromocytoma cells were purchased from Pasleur Institute (Tehran, Iran). Maltol (3-hydroxy-2-methyl-4-pyron), aluminum chloride (AlCl3.6H2O), 5,5′-dithiobis (2-nitrobenzoic acid) (DTNB) and sodium benzoate were bought from Merck Company (Darmstadt, Germany). Rhodamine 123 was obtained from Sigma-Aldrich Company (St. Louis, MO, USA). Also, fetal bovine serum, Dulbecco’s modified Eagle’s medium (DMEM), horse serum, trypsin, penicillin, and streptomycin were obtained from Gibco BRL Life Technologies (Grand Island, NY, USA).

Preparation of aluminum maltolate complex: Aluminum maltolate (Almal) was prepared according to a previous procedure [7, 19]. A stock solution (25 mM) of Almal was prepared and sterilized, using a 0.22 μM filter.

Cell culture & treatment: PC-12 cells were cultured in DMEM, containing 10% heat-inactivated horse serum, 5% fetal bovine serum, 100 U/mL penicillin and streptomycin. The cells were incubated at 37°C under 5% CO2 condition to reach 70-80% confluency. They were seeded overnight and then treated with Almal (250, 500, 750, or 1000 μM) or at various concentrations of sodium benzoate (0.125, 0.25, 0.5, 1, 1.5, 2, 2.5, or 3 mg/mL) in Phosphate Buffered Saline (PBS), and incubated at 37°C for 48hr. After analyzing the results of the cytotoxicity by 3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium (MTT) assay, Almal was selected for co-treatment at 500 μM concentration with sodium benzoate (0.125, 0.25, 0.5 or 1 mg/mL), and incubated for 48 hr.

Cell viability & apoptosis: After 48hr of treatment, MTT assay was used to determine the cell viability (10^4 cells/well). The medium was removed and 20 μL of MTT solution (5 mg/mL in PBS) was added, and incubated for another 4hr at 37°C. To dissolve the blue formazan product, 150 μL dimethylsulfoxide (DMSO) was added to each well. The absorbance was read at 570nm by an automated plate reader (BioTek, Bad Friedrichshall, Germany). Results were reported as percentages of cell viability relative to the untreated cells, as the control group. To determine cell apoptosis in the treated and control groups, we used Annexin V-FITC kit (Dako, catno: k235011). Briefly, after cell culture and treatment, cells were washed in cold PBS, suspended in 1mL ice cold binding buffer and added 5 μL Annexin V-FITC + 5 μL PI to 95 μL suspended cells, after gently mixing, incubated for 15 min in the dark. After adding 400 μL binding buffer, all tubes were analyzed with BD flow cytometer.

Measurement of Mitochondrial Membrane Potential (MMP): We used fluorescent dye Rhodamine-123 (Rh-123) for the measurement of MMP [7]. 10 μL Rh-123 was added to 2×10^5 cells and incubated for 30 min in the dark at 37°C. After washing with PBS, the fluorescence intensity was measured, using FLUO-star Omega multifunctional microplate reader (Munster, Germany). The wavelengths for excitation and emission were set at 485 and 525 nm, respectively [20].

Measurement of acetylcholinesterase activity: For measuring AChE activity, we used acethyl thiocholin as the substrate for AChE, as described by Ellman method [21]. Briefly, 3×10^6 cells were seeded in 6-well plates. After 48 hr of treatment, the media was removed and PBS added. Cells were then transferred to tubes for a 5-min centrifugation at 1500 rpm. Cell lysate was prepared by sonication, a 50 μL
cell lysate was added to 96-well plate plus 97 µL Ellman buffer (90 µL PBS + 5 µL DTNB + 2 µL acetylthiocholin). During the 15-min, absorption changes were recorded by an ELISA reader at 405nm. The protein content for each cell group was measured by the Bradford method [22] and the enzyme activity was determined.

**Statistical analysis:** The data from the triplicate, independent experiments are presented as Means±SEM. Data analysis was performed using one-way analysis of variance (ANOVA) followed by LSD post-hoc test on SPSS, v. 16. P values of <0.05 were considered as the statistical significance.

**Results**

**Effects on PC-12 cell line:** Based on our previous study on cell viability, we used 500 µM of Almal concentration, which is closest to LC50, and a co-treatment with non-cytotoxic concentration (0.125, 0.25 or 0.5 mg/mL) of sodium benzoate [19]. The co-treatment of Almal (500 µM) and sodium benzoate confirmed the protective effects of sodium benzoate.

![Figure 1. Effects of 48 hr of co-treatment with varying concentrations of sodium benzoate and Almal on cell viability.](image)

Panel A: Annexin V-PE and 7-AAD staining was represented by flow cytometry. Panel B: Q1 = necrotic; Q2 = late apoptotic cells; Q3: early stage apoptotic cells; Q4: normal viable cells.

Formaldehyde 3% used as a positive control. Data are expressed as the M means±SEM of triplicate, independent experiments.

Significant difference P < 0.05 (*=compared to the negative control group, #=compared to the Almal group). Almal=aluminum maltolate; SB, sodium benzoate.
benzoate at 0.25 and 0.5 mg/mL to be significant (P<0.05) as previously described [19].

**Effects on PC-12 Cell Apoptosis:** The flow cytometric analysis (Figures 1A & 1B) showed a decreasing cell viability to 66.37±0.96% of the counted cells, increasing percentage of apoptotic (30.2±9.7%) and necrotic (5.93±0.2%) cells after treatment with Almal (500 μM). In addition, co-treatment with sodium benzoate significantly increased cell viability and decreased apoptosis only at 0.5 mg/mL (P<0.05) without changing the number of necrotic cells compared to those for the Almal experiment.

**Effects on Mitochondrial Membrane Potential:** Our results did not show any significant decline neither in the MMP of PC-12 cells in Almal treatment vs. the control group, nor in the co-treatment of the cells with varying concentrations of sodium benzoate (Figure 2).

**Effects on Acetylcholinesterase Activity:** After treating PC-12 cells with Almal, the measured AChE activity showed a decline in the treated group compared to those in the control group. Moreover, co-treatment with sodium benzoate at 0.125, 0.25 or 0.5 mg/mL increased the AChE activity significantly, compared to those for the Almal group (P<0.05) (Figure 3).

**Figure 2.** Effects of 48 hr of co-treatment with varying concentrations of sodium benzoate and Almal

Mitochondrial Membrane Potential (MMP) was determined by Rhodamine 123. Data are expressed as the Mean±SD of triplicate, independent experiments. Almal=Aluminum maltolate; SB=Sodium benzoate.

**Figure 3.** Effects of 48 hr of co-treatment with varying concentrations of sodium benzoate and Almal

Acetyl cholinesterase activity (AChE) was determined by Ellman method. Data are expressed as the Mean±SD of triplicate independent experiments (#=compared to the Almal group). Almal=Aluminum maltolate; SB=Sodium benzoate.
Discussion

Since aluminum is increasingly used for numerous commercial purposes, such as healthcare, food packaging and agriculture, continued research is needed to explore its toxicity, and the association with neurodegenerative disorders [14]. Considering the varying effects of sodium benzoate and its potential role in the central nervous system, this study was conducted to investigate these effects. We investigated the effects of sodium benzoate on PC-12 cell line, with a focus on apoptosis, necrosis, mitochondrial membrane potential and acetylcholinesterase activity versus Almal toxicity.

Almal is a lipophilic compound, which is permeable through the cell membranes [23]. Following the metabolism of starch and sucrose, maltolate is produced in large quantities, which can bind to aluminum to produce Almal [24]. Studies have shown the protective effects of sodium benzoate in the management of cognitive disorders, such as AD, Parkinson’s Disease and multiple sclerosis [25-28]. Conversely, there are reports suggesting that sodium benzoate may have damaging effects on the mitochondrial functions [29]. Specifically, sodium benzoate may damage the mitochondrial DNA and the proteins involved in the electron transfer chains by generating free radicals [29]. In a study of sodium benzoate on HeLa cells and rat neurons, there were indications of apoptosis and disruption of calcium balance and mitochondrial membrane potentials [29]. In addition, previous studies have indicated that the oral administration of sodium benzoate induced anxiety, motor and memory impairments, increased brain oxidative stress and renal injury in vivo [30-33].

In PC-12 cell line, Almal induced cell death dose dependently, which is consistent with similar findings reported by a previous study [33]. At the concentrations of 0.015, 0.25, or 0.5 mg/ml, sodium benzoate had no deleterious effects on cell growth and the survival rate of PC-12 cells compared to the controls [7]. Therefore, these sodium benzoate concentrations were selected in the current study. This part of the experiments led to similar findings as those reported by a previous study [20]. The authors reported that sodium benzoate did not cause toxic effects on neurons up to a concentration of 1000 μg/ml [27]. Further, co-treatment of PC-12 cells with sodium benzoate and Almal showed that sodium benzoate (0.125-0.5 mg/mL) inhibited cell death due to the toxicity of Almal after 48 hr of incubation. In our previous study on PC-12 cells [19], sodium benzoate did not show protective effects against cell death at concentrations of 1-3 mg/mL, based on the MTT Assay. This was probably due to its toxicity at higher doses. The protective effect of sodium benzoate at 0.5 mg/mL against apoptosis induced by 48hr of exposure to Almal was significant (P<0.05). The cell death due to Almal was indeed apoptosis, as has also been reported by another study [34]. The data on the effects of aluminum and sodium benzoate did not suggest impairment in the Mitochondrial Membrane Potential (MMP). In another study that investigated the mechanism of induced apoptosis by aluminum, increases in the activity of Caspase 3 and P-53, and decreases in Bcl-2 and BAX factors were reported [24]. It appears that the mechanism of apoptosis induced by Almal or the inhibition of apoptosis by sodium benzoate is independent of changes in the mitochondrial membrane potential. The most protective mechanism of sodium benzoate is believed to be scavenging free radicals, such as hydroxyl groups [24].

Almal plays a significant role in the production of free radicals [28]. In addition, several studies have shown that the oxidative stress, induced by Almal, changes the lipid peroxidation and antioxidant enzymes activities [35, 36]. It has been suggested that the reduction in the mitochondrial membrane potential might not be necessary to induce apoptosis. Indeed, it may occur as a result of other signals to initiate the apoptosis pathway [37-39]. Therefore, it is believed that MMP impairment is not an early step in the initiation of apoptosis, as it might depend on cell types, other inducers and the type of fluorochrome marker used [40]. There are not many studies published on the effect of aluminum on mitochondrial membrane potential in mammalian cells, but recently one study suggested that aluminum reduces the membrane potential in lymphocytes [41].

Under physiological conditions, acetylcholinesterase is involved in the function of cholinergic neurons. Therefore, inhibition of acetylcholinesterase has been suggested as a therapeutic approach in the management of patients with Alzheimer’s disease [42]. The effects of sodium benzoate on acetylcholinesterase in the aluminum toxicity model and its role in the improvement of Alzheimer’s symptoms have previously been investigated [34]. The results suggest that the enzyme did not alter in PC-12 cells treated with either Almal or sodium benzoate. Further, this enzyme in patients with AD leads to strengthening of the acetylcholine messenger route and reduction of the cognitive impairment [43]. Based on the results of the current study, it can be concluded that sodium benzoate influences the activity of AChE and may have positive effects on the improvement of Alzheimer’s symptoms. In future studies, besides measuring the AChE activity, its levels in the CSF or blood or both should also be measured. Upon our literature search and to the best of our knowledge, there is no previous report on the effect of sodium benzoate on AChE activity in PC-12 cell line. Therefore, this study may be the first report in this context.

Due to limited laboratory resources, we could not measure cytochrome C activity to differentiate cell death derived from
mitochondrial versus non-mitochondrial dependent pathway, arising from the ROS levels and other oxidative stressors.

In addition to the measurement of cytochrome C activity, as stated above under the limitations, we recommend that future studies investigate the effect of sodium benzoate on the gene and protein expressions of mitochondrial respiratory chain complex. Besides measuring the AChE activity in future research, one should consider evaluating the AChE levels in the CSF or serum or both. Also, studying the rate of energy metabolism in similar cell lines and in animal models representing AD will help elucidate the protective effect of sodium benzoate in prevalent neurodegenerative disorders.

Conclusions

Based on the findings of this study, sodium benzoate exerts its protective effect against aluminum toxicity on cells through the inhibition of apoptosis. Both sodium benzoate and aluminum seem to impact similar targets for the AChE activity but with opposing outcomes. It also appears that the disruption of mitochondrial membrane potential has nothing to do with apoptosis induced by aluminum toxicity. Further, sodium benzoate is likely to relieve the symptoms of neurodegenerative diseases by lowering the incidence of neural cell death. The cellular and molecular mechanisms of this effect await future research.

Ethical Considerations

Compliance with ethical guidelines

This study was approved by the Ethics Committee of the University of Shiraz University of Medical Sciences (Ethical Code: IR.SUMS.REC.1394.S1116).

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Author’s contributions

All authors were equally contributed in preparing this article.

Conflict of interest

The authors declared no conflict of interest.

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