Protective Effects of Hydroalcoholic Extract of Nasturtium officinale

on Rat Blood Cells Exposed to Arsenic

Felor Zargari^{*1}, Amir Ghorbanihaghjo², Hossein Babaei³

Received: 27.01.2015

Accepted: 10.03.2015

ABSTRACT

Background: Arsenic is one of the most toxic metalloids. Anemia and leukopenia are common results of poisoning with arsenic, which may happen due to a direct hemolytic or cytotoxic effect on blood cells. The aim of this study was to examine the effects of hydroalcoholic extract of *Nasturtium officinale* on blood cells and antioxidant enzymes in rats exposed to sodium (meta)arsenite.

Methods: 32 Male Sprague Dawley rats were randomly divided into four groups; Group I (normal healthy rats), Group II (treated with 5.5mg/kg of body weight of NaAsO₂), Group III (treated with 500mg/kg of body weight of hydro-alcoholic extract of *N. officinale*), and Group IV (treated with group II and III supplementations). Blood samples were collected and red blood cell, white blood cell, hematocrit, hemoglobin, platelet, total protein and albumin levels and total antioxidant capacity were measured. Data was analyzed with Mann-Whitney U test.

Results: WBC, RBC and Hct were decreased in the rats exposed to NaAsO₂ (p<0.05). A significant increase was seen in RBC and Hct after treatment with the plant extract (p<0.05). There was no significant decrease in serum albumin and total protein in the groups exposed to NaAsO₂ compared to the group I, but NaAsO₂ decreased the total antioxidant capacity, significantly.

Conclusion: The *Nasturtium officinale* extract have protective effect on arsenic-induced damage of blood cells.

Keywords: Arsenic; Blood Cells; Nasturtium officinale; Oxidative Stress.

IJT 2015; 1331-1335

INTRODUCTION

Arsenic is one of the most toxic metalloids which is found in soil, water and air as a potential human carcinogen [1]. Many studies have confirmed the production of free radicals during arsenic metabolism in the cell [1-4]. It has been reported that exposure to sodium arsenite decreased the antioxidant enzymes, e.g. superoxid dismutase (SOD), catalase (CAT) and glutathion peroxidase (GPx) activity, significantly [5].

Anemia and leukopenia are common results of poisoning with arsenic which may

happen due to a direct hemolytic or cytotoxic effect on blood cells [6]. Trivalant arsenics are potent inhibitors of thioredoxin reductase (a NADPH-dependent flavoenzyme), so they inhibit the cellular responses to oxidative stress [7]. Defense system against free radical-induced oxidative stress includes enzymatic antioxidants (superoxide dismutase, catalase and gluthation peroxidase) and non-enzymatic antioxidants (vitamin C, vitamin E, carotenoids and flavonoids) [8].

In recent years, attentions have been paid toward medicinal plants. Phenolic compounds, particularly flavonoids, have antioxidant

^{1.} Department of Biology, Research branch, Islamic Azad University, Tehran, Iran.

^{2.} Department of Clinical Biochemistry, Tabriz University of Medical Sciences, Drug Applied Research Center, Tabriz, Iran.

^{3.} PhDof Pharmacology, Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

^{*} Corresponding Author: E-mail: felorzargari@marandiau.ac.ir

activities [9, 10]. *Nasturtium officinale* (watercress) from Brassicacea family is a perennial plant that thrives in cold water and is found in streams and contains vitamins A, C, E, gluconasturtine and some minerals, e.g. iron and phosphorus [11, 12]. Watercress is used as a home remedy for hypertension, cardiovascular disease and hyperglycemia [13].

The aim of this study was to examine the effects of hydroalcoholic extract of *Nasturtium officinale* on blood cells and antioxidant enzymes in rats exposed to sodium (meta)arsenite (NaAsO₂).

MATERIALS AND METHODS

Plant material and extraction

Aerial parts of *N. officinale* were gathered from Kaleybar City, East Azerbaijan Province, Iran, and are identified (NO. 711Tbz-fph) by an expert faculty of Tabriz University of Medical Sciences, Iran (Dr. Nazemiyeh). The samples were air-dried and powdered and were kept in a closed container at 8°C until extraction. 700g of the powder was extracted three times with the mixture of ethanol/water (4:1) at the room temperature overnight. The solvent was completely removed by a rotary evaporator (Heidolph; Germany) at 50°C. The final residue of the extract was used for tests.

Animals and experimental groups

32 Male Sprague Dawley rats weighing 180-200g (Iran; Central Animal House of Tabriz Medical School) were housed at $22\pm2^{\circ}$ C and $60\pm5\%$ relative humidity with a 12h light/dark cycle. They had access to water and normal diet *ad libitum*. The experimental protocol was approved by the medical ethics committee, and all animals received human care in compliance with the guidelines of Tabriz University of Medical Sciences. After the adaptation period (7 days), the rats were randomly divided into four groups (n=8); Group I (normal healthy rats), Group II (treated with 5.5mg/kg of body weight of NaAsO₂), Group III (treated with 500mg/kg of body weight of hydro-alcoholic extract of N. *officinale*), and Group IV (treated with group II and III supplementations). The groups were treated every day for 28 days with their supplementations and after the day 28, blood samples were collected.

Assessments

Hematological parameters: Whole blood samples were collected using EDTA as anticoagulants and were analyzed automatically with an Auto Hematology MINDRAY (BC-2800) for for red blood cell (RBC), white blood cells (WBC), hematocrit (Hct), hemoglobin (Hb), platelet (Plt).

Biochemical analysis: Total Antioxidant Capacity (TAC) was measured using Randox TAC status test (United Kingdum; Randox Laboratories Ltd.) in serum. ABTS (2,2-Azinodi-[3-ethylbenzthiazoline sulphate]) was incubated with a peroxidase (metmyoglobin) and H_2O_2 to produce the ABTS radical cation that has a relatively stable blue-green color at 600nm. Antioxidants suppress this color production to a degree which is propotional to their concentration. Serum albumin and total protein were measured by an autoanalyzer (Abbott ALCYON^{TM300}; USA) and enzymatic Kits (Pars Azmoon; Iran).

Statistical analysis

Data was inserted into SPSS 16 software and was analyzed with Mann-Whitney U test to compare the groups at the significance level p<0.05.

RESULTS

WBC, RBC and Hct were decreased in the rats exposed to NaAsO₂, significantly (p<0.05). A significant increase was seen in RBC and Hct after treatment with the plant extract (p<0.05). There was no significant decrease in serum albumin and total protein in the groups exposed to NaAsO₂ compared to the group I, but NaAsO₂ decreased the total antioxidant capacity, significantly (Table 1).

Blood Cells; Hb: Heamoglobin; Hct: Hematocrit; Plt: Platelet).				
Parameters	Group I	Group I	Group III	Group IV
Hematological				
WBC (10 ⁹ /l)	13.80 ± 2.10	10.45±2.40*	13.00 ± 3.20	11.18 ± 1.30
RBC $(10^{12}/l)$	8.38±0.30	7.11±0.60*	7.80 ± 0.50	8.20±0.30**
Hb (g/dl)	14.17 ± 0.50	13.54 ± 2.00	14.15 ± 0.50	13.60 ± 1.20
Hct (%)	44.90 ± 1.50	39.04±2.90*	43.53 ± 1.40	42.44±2.80**
Plt $(10^{9}/l)$	564.60 ± 59.70	678.70±77.90*	519.60 ± 95.00	663.30 ± 45.90
Biochemical				
Alb (g/dl)	2.91 ± 0.10	$2.90{\pm}0.1$	$3.08 \pm 0.20*$	3.39±0.20**
TP(g/dl)	$7.84{\pm}0.40$	$7.80{\pm}0.6$	$8.35\pm\!0.40*$	8.87±0.70**
TAC (mmol/dl)	0.89±0.13	$0.67 \pm 0.23*$	1.07±0.33**	1.15±0.37**

 Table 1. The average of hematological and biochemical parameters analysis of all 4 groups (Alb:

 Albumin; TP: Total Protein; TAC: Total Antioxidant Capacity; WBC: White Blood Cells; RBC: Red

 Blood Cells; Hb: Heamoglobin: Hct: Hematocrit; Plt: Platelet)

*p<0.05 compared to group I; **p<0.05 compared to group II

DISCUSSION

WBC, RBC and Hct were decreased in the rats exposed to sodium arsenite. The reduction of these hematological parameters might be due to the effect of arsenic on heme synthesis by inhibition of Delta-aminolevulinic acid dehydratase (ALAD), detoxification mechanism of arsenic (methylation of arsenic and depletion of gluthation). inhibition of pyruvate dehydrogenase and reduction of ATP and oxidative damage to blood cells.

Our results suggest that hydro-alcoholic extract of *N. officinale* may improve some hematological parameters. Many studies have reported that arsenic decreased WBC, RBC and hemoglobin [14-17]. Bone marrow depression in humans has been reported with high dose of arsenic [18]. 95-99% of the arsenic is located in erythrocytes after absorption, bound to the globin of hemoglobin and transported to the other parts of the body [19]. Rats differ from most mammalian species by accumulating dimethyl arsenic in erythrocytes probably by binding to a cysteine component of hemoglobin [20, 21].

Anemia and leukopenia may be due to hemolytic or cytotoxic effect of arsenic on the blood cells. The mechanism of hemolysis is depletion of intracellular GSH (gluthation) resulting in oxidation of sulfhydryl groups in the hemoglobin from ferrous to ferric in mice and rats [18, 22]. Erythrocytes may be susceptible to the oxidative damage due to the presence of heme iron, polyunsaturated fatty acids (PUFA) and oxygen, which may initiate the reactions that induce oxidative changes in RBC [23, 24]. Inhibition of pyruvate dehydrogenase (by binding of arsenic to dihydrolipoic acid, a pyruvate dehydrogenase cofactor, and inhibiting the conversion of pyruvate to acetyl coenzyme-A) or reduction of succinyl coenzyme-A decrease the production of ATP and may damage the cell slowly [18, 25, 26]. Antioxidant enzymes activity was decreased in rats exposed to sodium arsenite [27]. So sodium arsenite-induced oxidative stress and reduction of antioxidant enzymes activity and inhibition of heme synthesis may lead to the damage of blood cell and reduction of RBC, WBC and Hct in the present study.

Treatment of rats with polyphenols might have ameliorated the hematological system of rats exposed to lead [28]. Watercress may be able to improve antioxidant enzymes activity and some of hematological parameters due to the phenolic compounds or other compounds, e.g. isothiocyanates or beta-carotene.

Consumption of the plant extract with- (group IV) and without sodium arsenite (group III) showed a significant increase in serum levels of albumin, total protein and total antioxidant capacity. Tanju & Madhuri have reported that sodium arsenite decrease total protein and albumin in serum [29]. Increasing of serum protein and albumin has been reported with

consumption of ascorbic acid and plant antioxidants [29, 30]. Albumin plays an important role in transportation of some compounds in blood, so increasing of albumin in serum may help transporting of plant extract and sodium arsenite in blood. As these proteins are produced in the liver, any changes in the concentration of serum proteins and albumin indicate a change in the normal function of liver [30]. Plant extract has been reported to decrease the lipid peroxidation and aminotransferase activity and increase the status of antioxidant enzymes in liver [27]. So, increasing the total protein of serum is thought to be associated with hepato-protective effects of plant extract and improving of proteins synthesis in liver or immune responses [31]. Asadi et al. have indicated that watercress enhances globulin levels and improves the immune system [31]. Increasing of the total antioxidant capacity might be due to the effects of substances such as vitamin C, vitamin E, carotenoieds (e.g. βcarotene) or superoxide scavenging activity (e.g. quercetin) of flavonoids in the plant extract.

Further studies are needed to determine the possible mechanisms of action and specific bioactive compounds in *Nasturtium officinale*. It is recommended that long-term animal studies on depending use of other oxidants, the dose and route of arsenic exposure to evaluate the effects of the extract be carried out.

CONCLUSION

The *Nasturtium officinale* extract have protective effect on arsenic-induced damage of blood cells and can improve hematological parameters and total antioxidant capacity.

ACKNOWLEDGMENT

We wish to thank the director of Drug Applied Research Center of Tabriz University of Medical Sciences for supporting this study.

REFERENCES

 Blessing Ebele O. Mechanisms of arsenic toxicity and carcinogenesis. African J Bio Res. 2009;3(5):232-7.

- Singh S, Rana SV. Amelioration of arsenic toxicity by L-Ascorbic acid in laboratory rat. J Environ Biol. 2007;28(Suppl 2):377-84.
- 3. Saxena PN, Anand S, Saxena N, Bajaj P. Effect of arsenic trioxide on renal functions and its modulation by Curcuma aromatica leaf extract in albino rat. J Environ Biol. 2009;30(4):527-31.
- 4. Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chem Biol Interact. 2006;160(1):1-40.
- Ercal N, Gurer-Orhan H, Aykin-Burns N. Toxic metals and oxidative stress part I: Mechanisms involved in metal induced oxidative damage. Curr Top Med Chem. 2001;1(6):529-39.
- Kyle RA, Pease GL. Hematologic aspects of arsenic intoxication. New Engl J Med. 1965;273:18-23.
- Lin S, Del-Razo LM, Styblo M, Wang C, Cullen WR, Thomas DJ. Arsenicals inhibit thioredoxin reductase in cultured rat hepatocytes. Chem Res Toxicol. 2001;14(3):305-11.
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol. 2007;39(1):44-84.
- Hollman PCH. Absorption, bioavailability, and metabolism of flavonoids. Pharma Biol. 2004; 4(Suppl):74-83.
- Pool-Zobel BL, Bub A, Wollowski I, Rechkemmer G. Consumption of vegetables reduces genetic damage in humans: First results of a human intervention trial with carotenoid-rich foods. Carcinogenesis. 1997;18(9):1847-50.
- Soengas P, Sotelo T, Velasco P, Cartea EM. Antioxidant properties of Brassica vegetables. Functional Plant Sci Biotechnol. 2011;5(Special Issue 2):43-55.
- 12. Gill CI, Haldar S, Boyd LA, Bennett R, Whiteford J, Butler M, et al. Watercress supplementation in diet reduces lymphocyte DNA damage and alters blood antioxidant status in healthy adults. Am J Clin Nutr. 2007;85(2):504-10.
- 13. Zargari A. Medicinal plants. Tehran: Tehran University Press; 1999. pp. 199-204.
- 14. Gyasi SF, Awuah E, Larbi JA, Koffuor GA, Osei OA. Clinical, hematological and histopathological responses to arsenic toxicity in ICR mice using arsenic levels synonymous to buruli ulcer endemic communities in the amansie west district of Ghana. Eur J Exp Biol. 2012;2(3):683-89.

- 15. Ferzand R, Gadahi JA, Saleha S, Ali Q. Histological and hematological disturbance caused by arsenic toxicity in mice model. Pak J Biol Sci. 2008;11(11):1405-13.
- 16. Flora SJ, Chouhan S, Kannan GM, Mittal M, Swarnkar H. Combined administration of taurin and monoisoamyl DMSA protects arsenic induced oxidative injury in rats. Oxid Med Cell Longev. 2008;1(1):39-45.
- 17. Jain A, Yadav A, Bozhkov AI, Padalko VI, Flora SJ. Therapeutic efficacy of silymarin and naringenin in reducing arsenic-induced hepatic damage in young rats. Ecotoxicol Environ Saf. 2011;74(4):607-14.
- Saha JC, Dikshit AK, Bandyopadhyay M, Saha KC. A review of arsenic poisoning and its effects on human health. Crit Rev Environ Sci Technol. 1999;29(3):281-313.
- 19. Valee L, Ulmer DD, Wacker WEC. Arsenic toxicity and biochemistry. Arch Ind Heath. 1960;21:132-51.
- 20. Lu M, Wang H, Li XF, Arnold LL, Cohen SM, Le XC. Binding of dimethylarsinous acid to cys-13alpha of rat hemoglobin is responsible for the retention of arsenic in rat blood. Chem Res Toxicol. 2007;20(1):27-37.
- Vahter M. Biotransformation of trivalent and pentavalent inorganic arsenic in mice and rats. Environ Res. 1981;25(2):286-93.
- 22. Lerman BB, Ali N, Green D. Megaloblastic, dyserythropoietic anemia following arsenic ingestion. Ann Clin Lab Sci. 1980;10(6):515-7.
- 23. Flora SJ, Bhadauria S, Kannan GM, Singh N. Arsenic induced oxidative stress and the role of antioxidant supplementation during chelation: a review. J Environ Biol. 2007;28(2):333-47.
- 24. Szymanska-Chabowska A, Antonowicz-Juchniewicz J, Andrzejak R. Some aspects of

arsenic toxicity and carcinogenicity in living organism with special regard to its influence on cardiovascular system, blood and bone marrow. Int J Occup Med Environ Health. 2002;15(2):101-16.

- 25. Reichl FX, Szinicz L, Kreppel H, Forth W. Effect of arsenic on carbohydrate metabolism after single or repeated injection in guinea pigs. Arch Toxicol. 1988;62(6):473-5.
- 26. Hughes MF. Arsenic toxicity and potential mechanisms of action. Toxicol Lett. 2002;133(1):1-16.
- 27. Zargari F, Ghorbanihaghjo A, Babaei H, Farajnia S, Roodbari NH. The effect of hydroalcoholic extract of Nasturtium officinale R. Br on antioxidant status and DNA damage in liver and kidney rats exposed to arsenic. Med J Tabriz Uni Med Sci. 2014;36(3):44.
- 28. Aksu DS, Didin M, Kayikci F. The protective role of polyphenols on blood cells in rats exposed to lead. Rev Roma de Med de Lab. 2012;20(3-4):47-57.
- 29. Tanju S, Madhuri D. Arsenic induced oxidative stress, hemato-biochemical and histological changes in liver and protective effect of moringa leaf powder and ascorbic acid in broiler chicken. J Chem Pharm Res. 2013;5(2):112-6.
- 30. Udenze ECC, Braide VB, Okwesilieze CN, Akuodor GC, Odey MO. The effects of gavage treatment with Garcinia kola seeds on biochemical markers of liver functionality in diabetic rats. Ann Biol Res. 2012;3(9):4601-8.
- 31. Asadi MS, Miirvaghefei AR, Nematollahi MA, Banaee M, Ahmadi K. Effects of watercress (Nasturtium Officinale) extract on selected immunological parameters of rainbow trout (Oncorhynchus mykiss). Open Vet J. 2012;2:32-9.